

5.2.12 Miscellaneous Diseases of Molluscs

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This section is included to provide brief information on a variety of types of diseases that the diagnostician may hear of or be asked for information on. These diseases are not given detailed treatment for one of the following reasons: (1) they are essentially opportunistic diseases that, while important in shellfish husbandry, are not regarded to have any significance in the geographic transport of shellfish, or (2) there is insufficient information on the significance of the diseases to individuals or populations of shellfish or on their etiology and diagnosis, or (3) they are diseases of species that have limited commercial importance with the result that less is known about their geographic distribution and significance (e.g. haplosporidian infections in the gaper clam or in boring bivalves). It must also be noted that some parasites and infectious diseases are completely omitted but will likely be included in subsequent editions as their significance to aquaculture and resource management becomes more widely known. The number of reports of significant mollusc diseases is increasing rapidly. Due to the inevitable lag time in the publication of a diagnostic guide such as this one, the most recently reported diseases will not be included.

A. Rickettsia- and Chlamydia-like Diseases of Bivalve Molluscs

Rickettsia-like and chlamydia-like organisms are some of the most common histopathological observations in bivalves. There has been no definitive evidence that these are linked in terms of pathogenicity to the morphologically similar human and mammalian pathogens. They most typically are observed in epithelial cells of the digestive gland or gill (Figure 1 and Figure 2). In a few cases they are reported to be associated with mortalities of the host. However, this has not yet been definitively demonstrated and there are many examples of apparently benign infections. Chlamydia-like organisms have been reported in the bay scallop *Argopecten irradians*, the Portuguese oyster *Crassostrea angulata*, and the hard clam *Mercenaria mercenaria*. Rickettsia-like organisms have been reported in the Pacific oyster *Crassostrea gigas*, the Eastern oyster *Crassostrea virginica*, *Donax trunculus*, *Mercenaria mercenaria*, the soft shell clam *Mya arenaria*, the sea scallop *Placopecten magellanicus*, the Pacific razor clam *Siliqua patula*, the thin tellin *Tellina tenuis*, the manila clam *Tapes philippinarum*, the Japanese scallop *Patinopecten yessoensis*, the European flat oyster *Ostrea edulis*, and the Palourde clam *Ruditapes philippinarum*. The bacterial forms are usually distinguishable at the light microscope level but in some cases, as with the chlamydia-like structure in the digestive epithelium of *Mercenaria mercenaria*, the entire intracytoplasmic inclusion is referred to as an amorphous blue-body.



Figure 1. Rickettsia-like organisms in the branchial epithelium of the Japanese scallop (arrows) *Patinopecten yessoensis*, 580X. From Elston 1986, with permission.

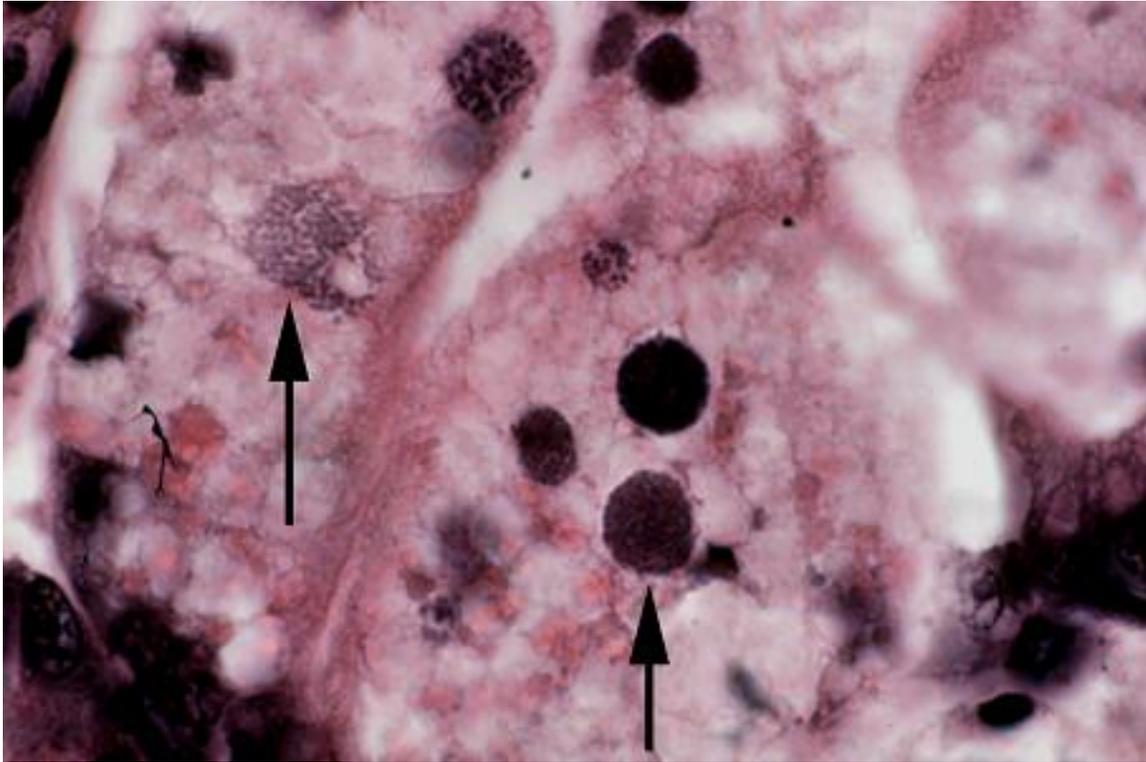


Figure 2. Rickettsia-like organisms in the digestive tubular epithelium of the Pacific razor clam *Siliqua patula* (examples at arrows), 880X.

B. Malpeque Bay Disease

This is a widely known but poorly understood disease that caused severe mortalities in American oysters *Crassostrea virginica* in Malpeque Bay along the Canadian maritime province of Prince Edward Island starting in 1915 and continuing through the 1930s. The geographically progressive nature of the disease, which was first observed a year after substantial plantings of seed oysters imported from the United States, is considered evidence for an infectious etiology of the disease. Over 90% of original stocks are reported to have succumbed to the disease, characterized by visceral shrinkage, a translucent quality, reduced growth, and failure to spawn. The etiology has never been determined with certainty.

C. Vibriosis of Larval and Juvenile Molluscs

This is regarded as an opportunistic disease of intensively cultured molluscs. Although several species including *Vibrio tubiashi*, *Vibrio anguillarum*, and *Vibrio alginolyticus* are reported associated with the diseases, it is likely that other poorly characterized marine vibrios are also important in the disease in some locations and circumstances. Usually the bacteria can be reduced or eliminated in the husbandry systems by hygienic or water treatment modifications. Vibriosis is more likely to occur in the warmest months of the year. Pathogenesis based on both exotoxins and direct invasion of tissues has been reported.

D. Gill Disease of Portuguese Oysters *Crassostrea angulata*

Severe mortalities are attributed to this disease in the Ile de Oleron and Arcachon regions of France in 1967-68 and 1970-73. It has also been reported in Great Britain. The disease has effectively destroyed *Crassostrea angulata* as a cultured oyster. Subsequently, and apparently as a result of this disease, the Pacific oyster *Crassostrea gigas* has replaced the culture of the Portuguese oyster in France. In the active stages of this disease, yellow spots appear on the gill. These spots progress and their centers become brown, necrotic, and disintegrate, leaving a perforation or an indentation on the gill. In the inactive or remissive state, occurring in some individuals, the perforations or indentations occur without the necrotic tissue. Yellow or green pustules may occur on the mantle or adductor muscle. Several etiologies have been proposed but none definitively proven. Iridovirus-like particles have been observed in the gill epithelium of some oysters affected with the disease.

E. Hinge Ligament Disease of Juvenile Bivalve Molluscs

This is a disease that was discovered in a variety of cultured juvenile bivalve molluscs up to about 1 cm in shell height. It is associated with substantial mortalities. Gliding or cytophaga-like bacteria invade and erode the proteinaceous hinge ligament, often resulting in its complete liquefaction and attendant dysfunction (Figure 3 and Figure 4). A portal of entry to the soft tissues may thus be opened, but this point has not been definitively established. The frequency of observation of this phenomenon in husbanded juvenile molluscs suggests that it is a serious problem. Morphologically similar lesions have been observed in several species in many locations on both coasts of North America suggesting a common or family of similar etiological agents. The presumptive causative organism can be cultivated by careful excision of the ligament and dilution on sea-water cytophaga agar (see references).

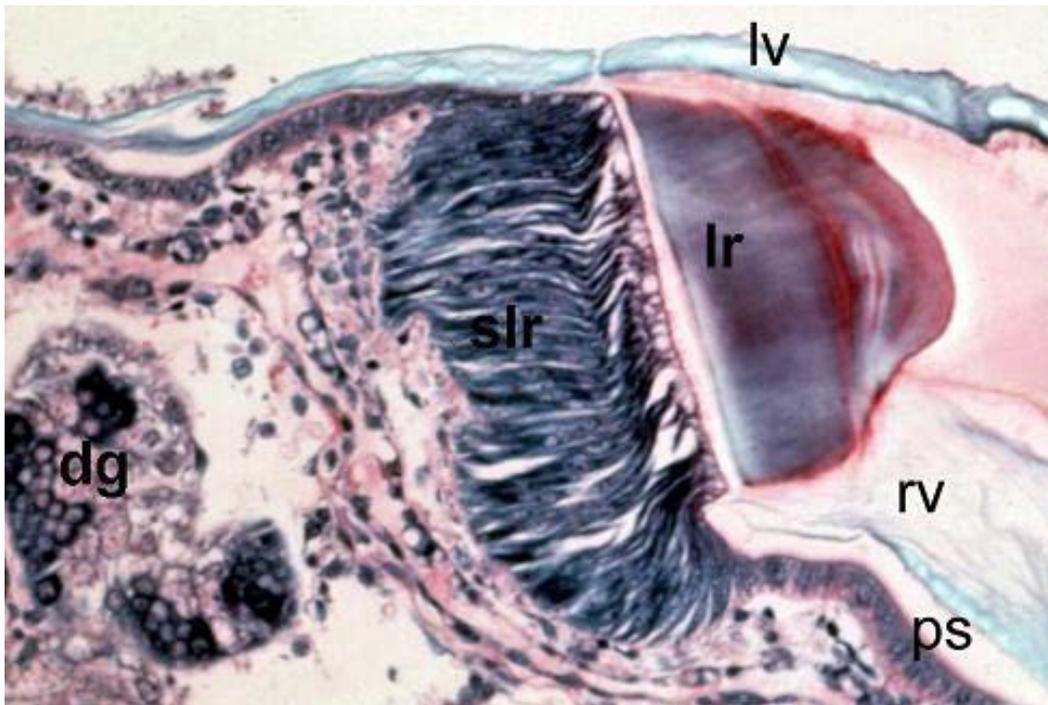


Figure 3. Normal hinge ligament of oyster showing the intact resilium (lr) and other adjacent organs (dg, digestive gland; slr, sub-ligamental ridge; lv, rv, left and right valves; ps, pallial space), 160X From Dungan and Elston 1988, with permission.

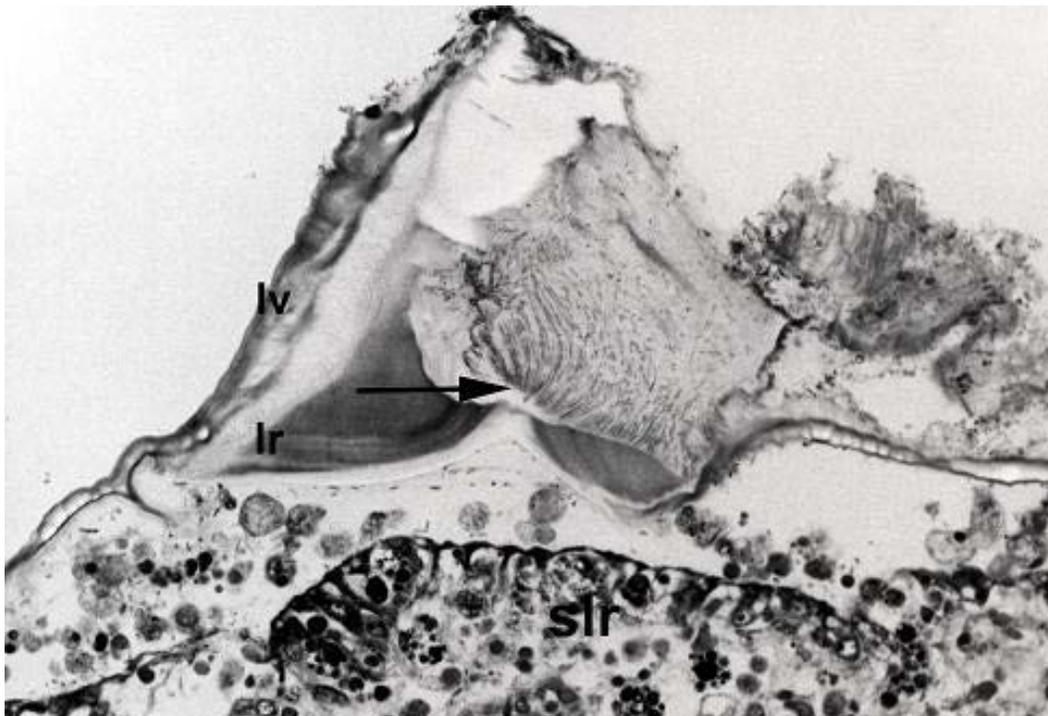


Figure 4. Hinge ligament eroded with cytophaga-like bacteria. The bacteria appear as fibers oriented at right angles to the eroding resilium margin (arrow). The sub-ligamental ridge appears necrotic and is separated from the internal resilium surface by a wide pallial space containing bacteria and rounded host cells. Organ identifiers and magnification same as Figure 3. From Dungan and Elston 1988, with permission.

References

- Armstrong, D. A., and J. L. Armstrong. 1973. A haplosporidan infection in gaper clams, *Tresus capax* (Gould), from Yaquina Bay, Oregon. *Proceedings of the National Shellfisheries Association* 64:68-72.
- Caty, X. 1969. Note preliminaire sur la presence de proliferations observees sur les huitres atteintes de la maladie des branchies. *Revue Travaux de l'Institut des Peches Maritimes* 33:167-170.
- Comps, M. 1982. Etude morphologique d'une infection rickettsienne de la palourde *Ruditapes philippinarum* Adam and Reeves. *Revue Travaux de l'Institut des Peches Maritimes* 46:141-145.
- Comps, M. 1983. Infections rickettsiennes chez les mollusques bivalves des cotes francaises. *Rapports et Proces-Verbaux des Reunions - Conseil International pour l'Exploration de la Mer* 182:134-136.
- Comps, M., J. P. Deltreil, and C. Vago. 1979. Un microorganisme de type rickettsienne chez l'Huitre portugaise *Crassostrea angulata* Lmk. *Comptes Rendu Academie des Science Paris*, 289, Series D:169-171.
- Comps, M., and R. Raimbault. 1978. Infection rickettsienne de la glande digestive de *Donax trunculus* Linne. *Science et Peche Bulletin de l'Institut des Peches Maritimes* 281.
- Comps, M., J-R Bonami, and C. Vago. 1977. Pathologie des invertebres-infection virale associee a des mortalities chez l'Huitre *Crassostrea gigas* Thunberg. *Comptes Rendus Academie des Science Paris*, 285 D:1139-1140.
- Dungan, C. F., and R. A. Elston. 1988. Histopathological and ultrastructural characteristics of bacterial destruction of the hinge ligaments of cultured juvenile Pacific oysters, *Crassostrea gigas*. *Aquaculture* 72:1-14.
- Dungan, C. F., R. A. Elston, and M. H. Schiewe. 1989. Evidence for colonization and destruction in hinge ligaments of cultured juvenile Pacific oysters (*Crassostrea gigas*) by cytophaga-like bacteria. *Applied and Environmental Microbiology* 55:1128-1135.
- Elston, R. A. 1986. An intranuclear pathogen [nuclear inclusion X (NIX)] associated with massive mortalities of the Pacific razor clam, *Siliqua patula*. *Journal of Invertebrate Pathology* 47:93-104.
- Elston, R. A. 1986. Occurrence of branchial rickettsiales-like infections in two bivalve molluscs, *Tapes japonica* and *Patinopecten yessoensis*, with comments on their significance. *Journal of Fish Diseases* 9:69-71.
- Farley, C. A., W. G. Banfield, G. Kasnic, Jr., and W. S. Foster. 1972. Oyster herpes-type virus. *Science* 178:759-760.
- Gulka, G., P. W. Chang, and K. A. Marti. 1983. Prokaryotic infection associated with a mass mortality of the sea scallop *Placopecten magellanicus*. *Journal of Fish Diseases* 6:355-364.
- Harshbarger, J. C., S. C. Chang, and S. V. Otto. 1977. Chlamydiae (with phages), mycoplasmas, and rickettsia in Chesapeake Bay bivalves. *Science* 196:666-668.

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- Hillman, R. E. 1978. The occurrence of *Minchina sp.* (Haplosporida, Haplosporidiidae) in species of the molluscan borer, *Teredo*, from Barnegat Bay, New Jersey. *Journal of Invertebrate Pathology* 31:265-266.
- Katkansky, S. C., A. K. Sparks, and K. K. Chew 1967. Distribution and effects of the endoparasitic copepod, *Mytilicola orientalis*, on the Pacific oyster, *Crassostrea gigas*, on the Pacific coast. *Proceedings of the National Shellfisheries Association* 57:50-58.
- Katkansky, S. C., and R. W. Warner. 1970a. The occurrence of a haplosporidan in Tomales Bay, California. *Journal of Invertebrate Pathology* 16:144.
- Katkansky, S. C., and R. W. Warner. 1970b. Sporulation of a haplosporidan in a Pacific oyster (*Crassostrea gigas*) in Humboldt Bay, California. *Journal of the Fisheries Research Board of Canada* 27:1320-1321.
- Meyers, T. R. 1979. Preliminary studies on a chlamydial agent in the digestive diverticular epithelium of hard clams *Mercenaria mercenaria* (L.) from Great South Bay, New York. *Journal of Fish Diseases* 2:179-189.
- Morrison, C., and G. Shum. 1982. Chlamydia-like organisms in the digestive diverticula of the bay scallop, *Argopecten irradians* (Lmk). *Journal of Fish Diseases* 5:173-184.
- Needler, A. W. H., and R. R. Logie. 1947. Serious Mortalities in Prince Edward Island Oysters Caused by a Contagious Disease. Pages 73-89 in *Transaction of the Royal Society of Canada. Volume XLI: Series III: Section V. May 1947.*
- Perkins, F. O., and P. H. Wolf. 1976. Fine structure of *Marteilia sydneyi* haplosporidan pathogen of Australian oyster. *Journal of Parasitology* 62:528-538.
- Taylor, R. T. 1966. *Haplosporidium tumefacientis* sp. n., the etiologic agent of a disease of the California sea mussel, *Mytilus californianus* Conrad. *Journal of Invertebrate Pathology* 8:109-121.